



# 4<sup>th</sup> MEETING ON INNOVATIVE IMMUNOTHERAPIES FOR LYMPHOID MALIGNANCIES

Presidents  
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## CAR-T for HL: Are we catching up?

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MILANO, STARHOTELS ROSA GRAND  
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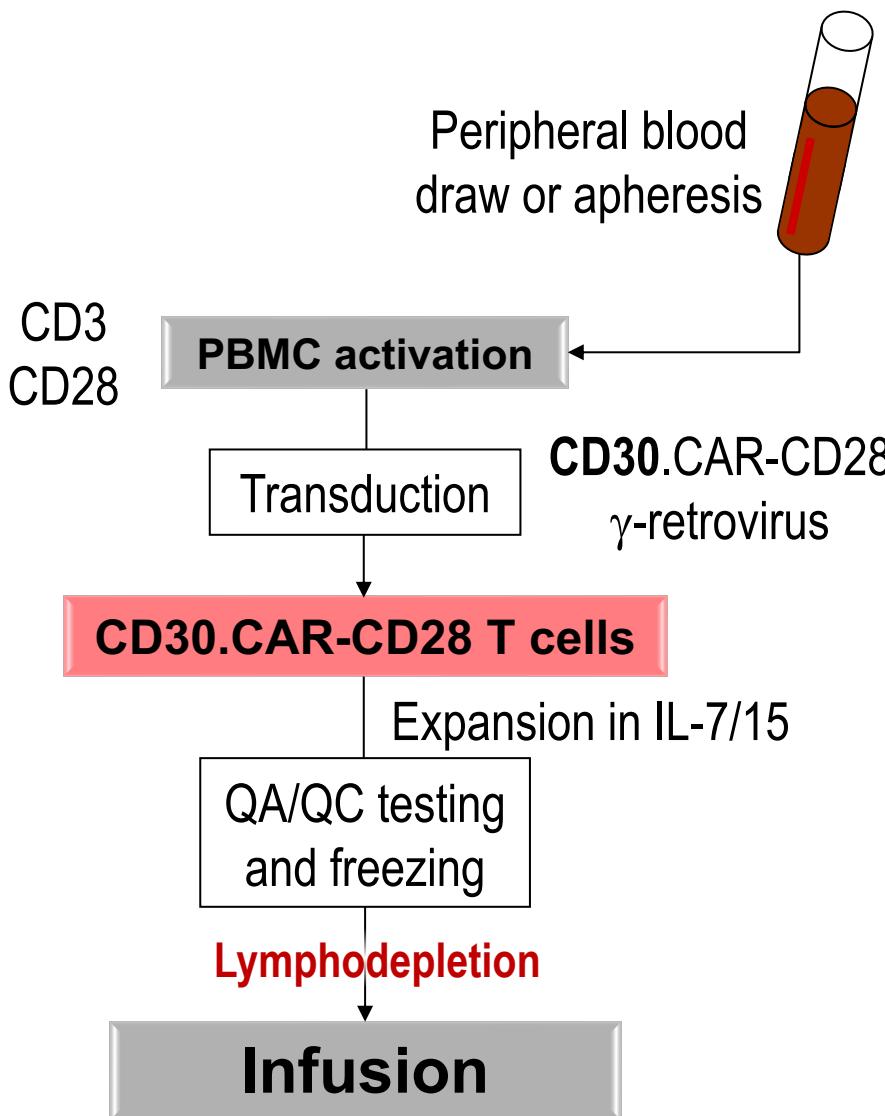
## Disclosures of Carlos A. Ramos

Company name	Research support	Employee	Consultant	Stockholder	Speakers bureau	Advisory board	Other
Tessa Therapeutics	x						
Athenex, Inc.	x						

# Targeting CD30 with a CAR

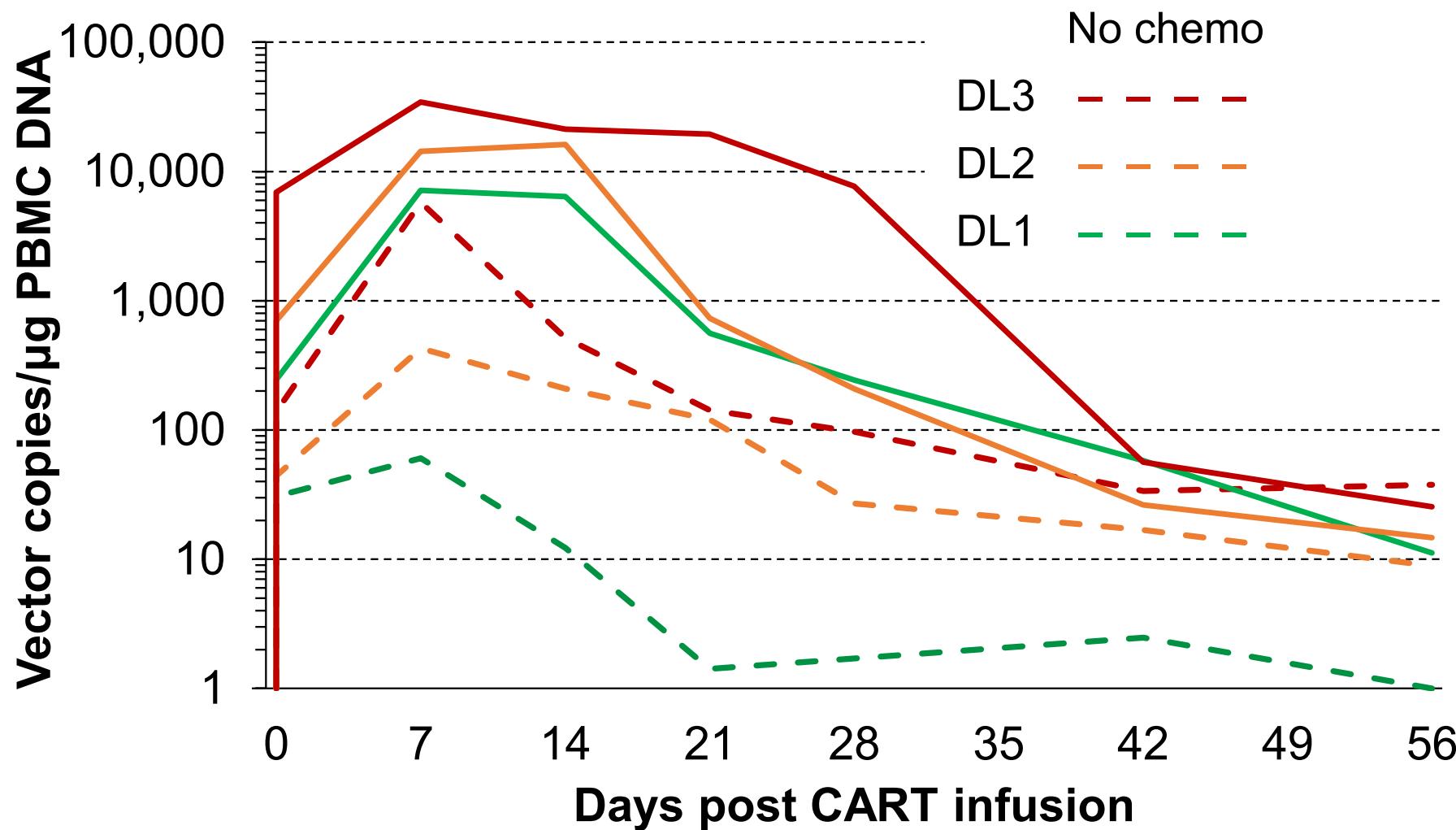
- CD19-specific (and BCMA) CAR-T cells are highly successful against B-cell NHL and ALL (and myeloma)
- Adequate targets for other disorders have been more difficult to define
- CD30 has been validated as an immune target (e.g. brentuximab vedotin)
- A CD30-specific CAR (CD30.CAR) has activity in pre-clinical models of HL (Hombach, Ca Res 1998; Savoldo, Blood 2007)

# ATLAS (UNC) & RELY-30 (BCM) trials



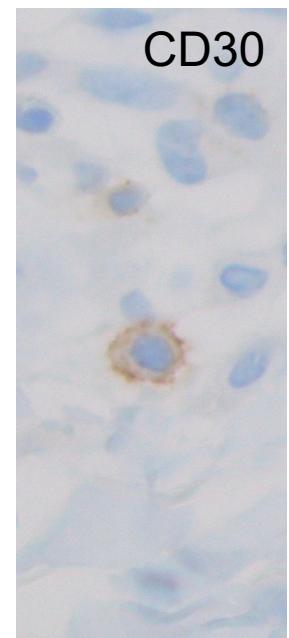
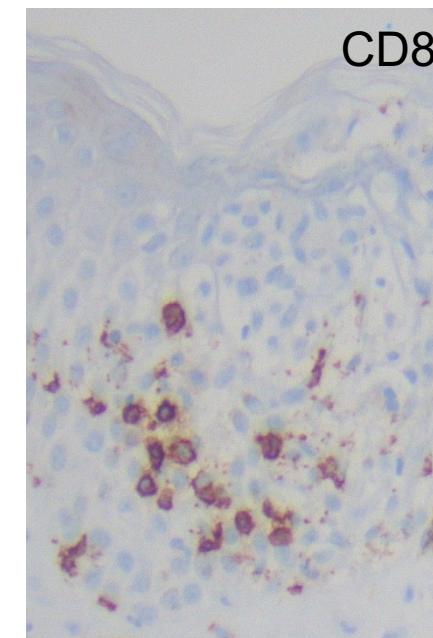
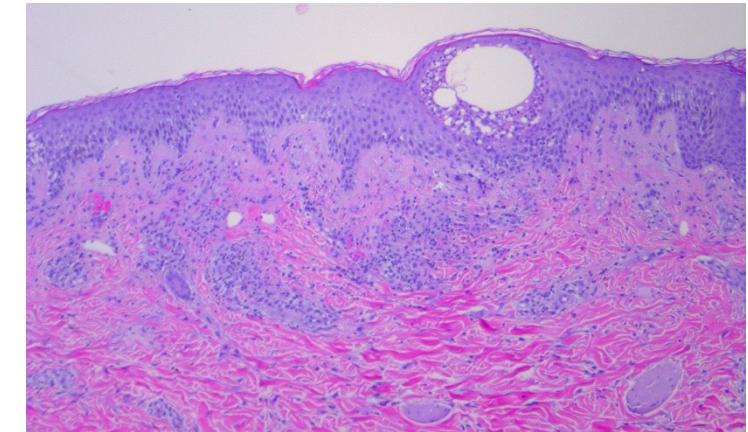
- 41 enrollments
- Gender
  - 13 F, 28 M
- Diagnoses
  - Hodgkin lymphoma (41)
    - Nodular sclerosis (32)
    - Mixed cellularity (4)
    - “NOS” (5)
- Median age 35 yrs (range 17-69)
- Median 7 prior treatments (range 2-23)
  - PD-1 inhibitor (34), brentuximab vedotin (38), HDT/ASCT (32), allo-SCT (10)

# CD30.CART expansion is increased by lymphodepleting chemotherapy



# Autologous CD30.CART main toxicities

- No neurotoxicity
- CRS in 10 pts
  - all grade 1
  - all resolved spontaneously
- Rash in 20 pts
  - all resolved spontaneously
  - 3 baseline rashes



Patient B9

# Grade 3 or higher toxicities

Toxicity (N= 42)	Grade 3/4 N (%)	Not resolved >28 d N (%)	Not resolved >3 mo N (%)
Lymphopenia	42 (100)	-	-
Neutropenia	20 (48)	4 (10)	0
Thrombocytopenia	11 (26)	10 (24)	4 (10)
Anemia	5 (12)	0	0
Pneumonia	1 (2)	-	-
Hypoalbuminemia	3 (7)	-	-
Hyponatremia	2 (5)	-	-

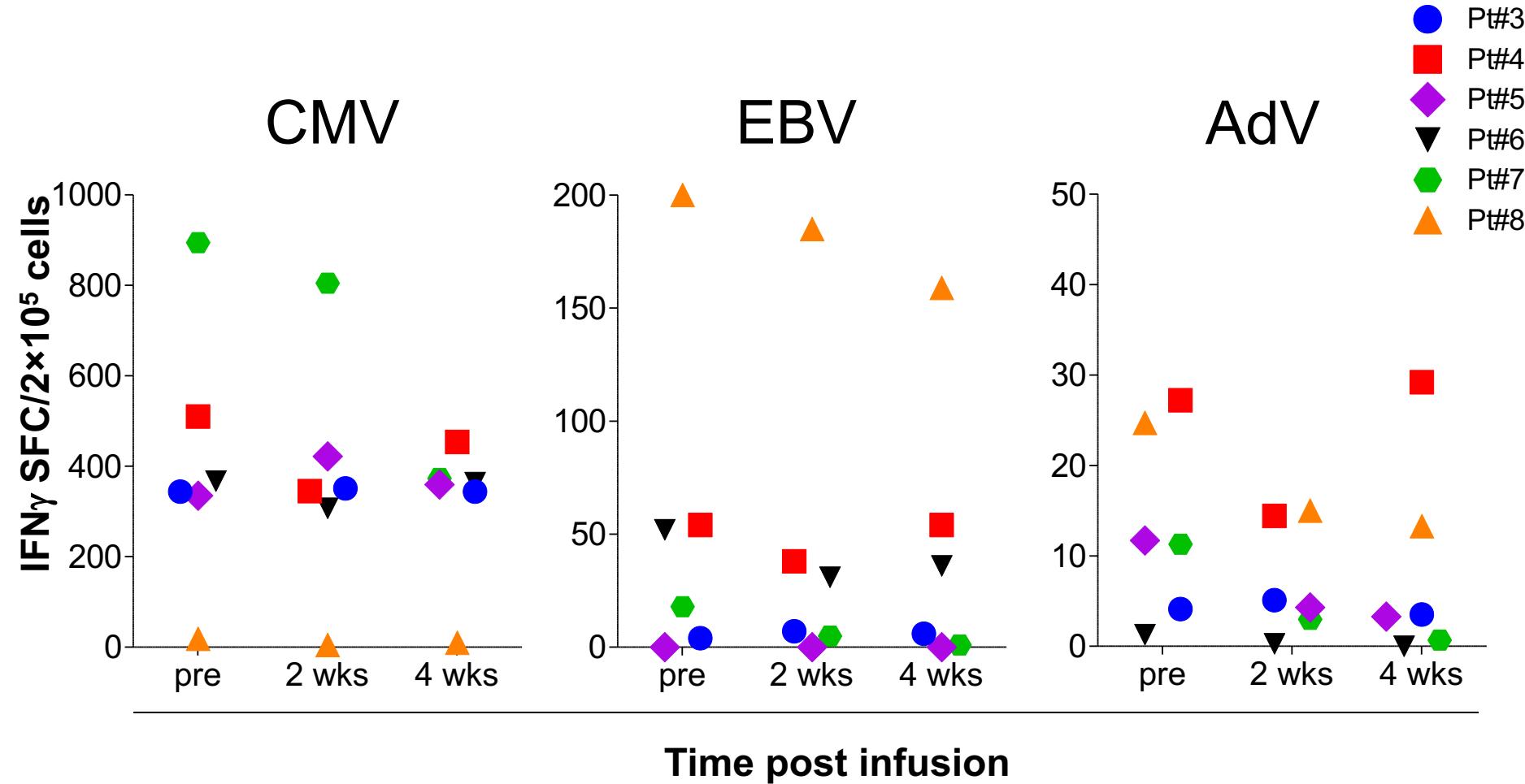
# Other potential concerns related to CD30 targeting

- CD30 is preferentially and/or constitutively expressed by Th2 or Tc2 cells
- CD30 is expressed transiently by activated T cells after exposure to cognate antigen

⇒ Need to ensure that CD30.CAR-T cells do not eliminate activated (viral) antigen-specific T cells *in vivo*:

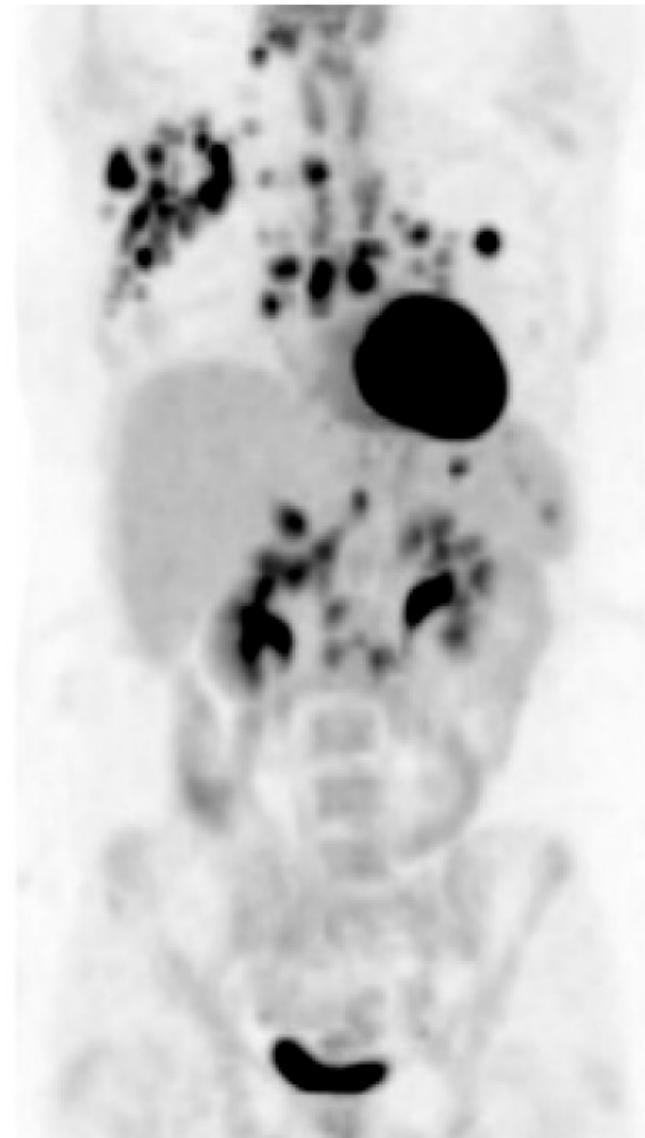
- pre and post infusion virus-specific immune response monitoring

# Viral immunity is not compromised

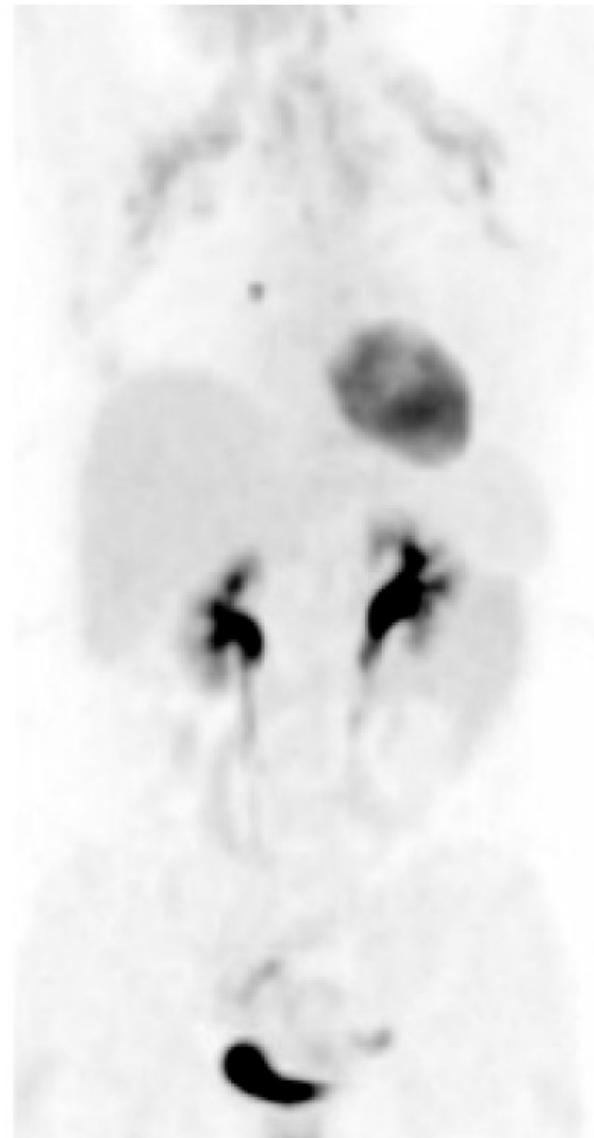


# Response to autologous CD30.CART

Pre-  
infusion

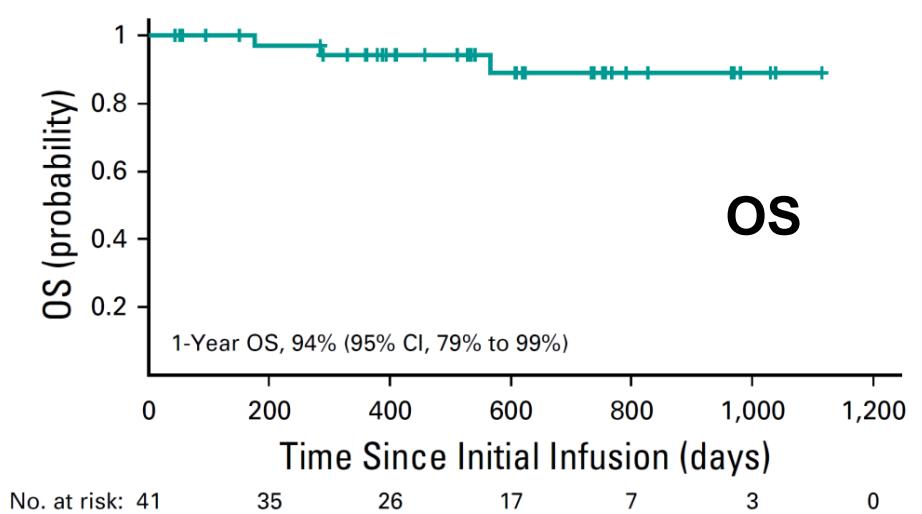
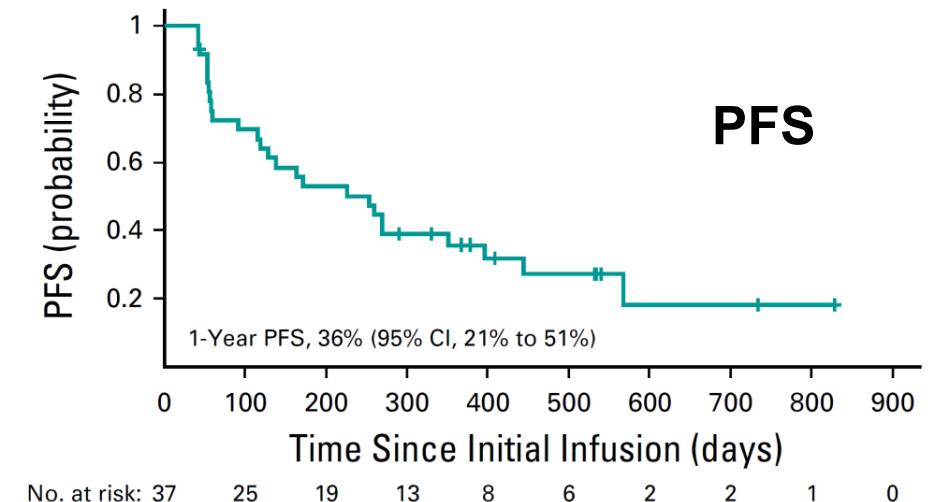
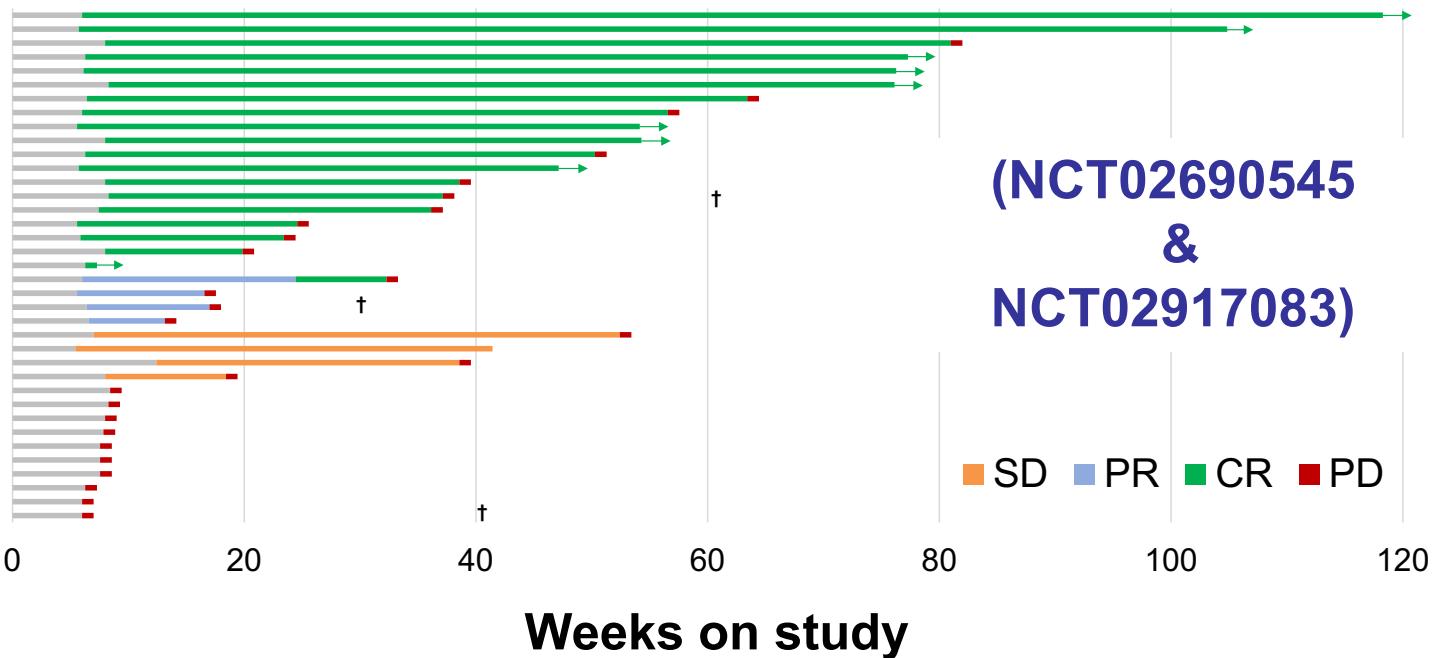


6 wks  
post-  
infusion



# Autologous CD30.CAR-T cells in HL (BCM/UNC)

- With optimal lymphodepletion:
  - 72% overall response rate
  - 59% complete responses



(Ramos, Grover *et al.*, J Clin Oncol 2020)

# CHARIOT (NCT04268706) trial

## Study Population

Patients with R/R cHL:

- 12-75 years old
- Failed  $\geq 3$  lines of therapy including:
  - Chemotherapy
  - Brentuximab vedotin, @ and
  - PD-1 inhibitor@

May have received an autologous or allogeneic stem cell transplant

## Study Treatment

(Pilot: n =  $>12$ ,  
Pivotal: n = 82)

### LD (3 days)\*

- Fludarabine 30 mg/m<sup>2</sup>/day
- Bendamustine 70 mg/m<sup>2</sup>/day

### CD30.CAR-T<sup>#</sup>

Allowable dose range:  
 $2.0-2.7 \times 10^8$  cells/m<sup>2</sup>

## Endpoints

### Primary

- Pilot: Safety
- Pivotal: ORR

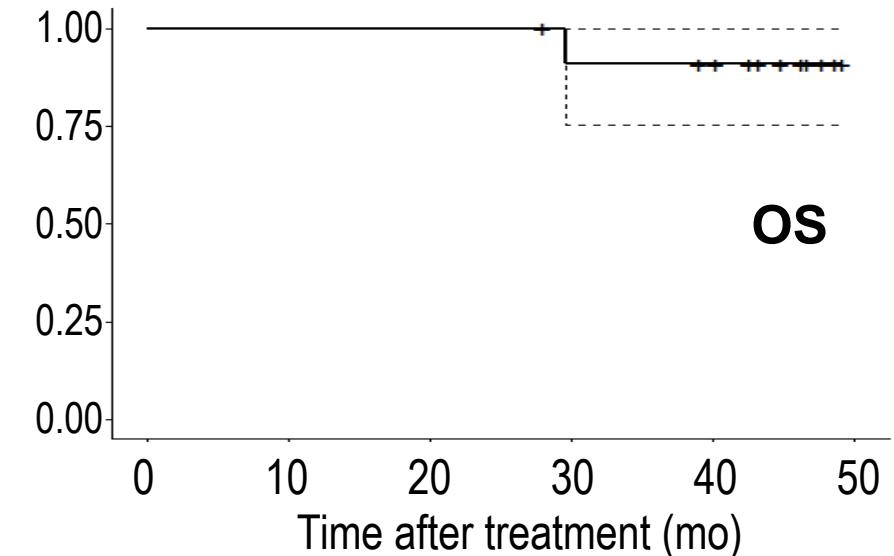
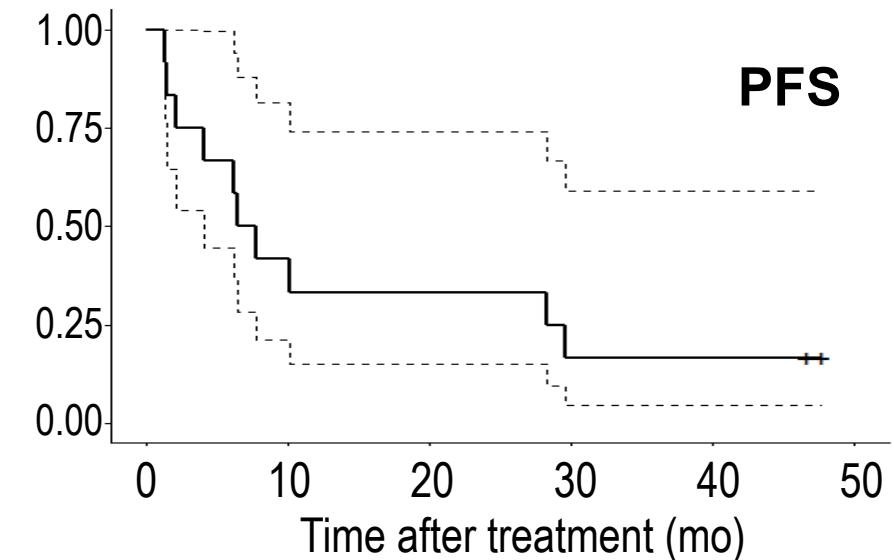
### Secondary

- Pilot:  
ORR, DOR,  
PFS, OS, HRQoL
- Pivotal:  
Safety, DOR,  
PFS, OS, HRQoL

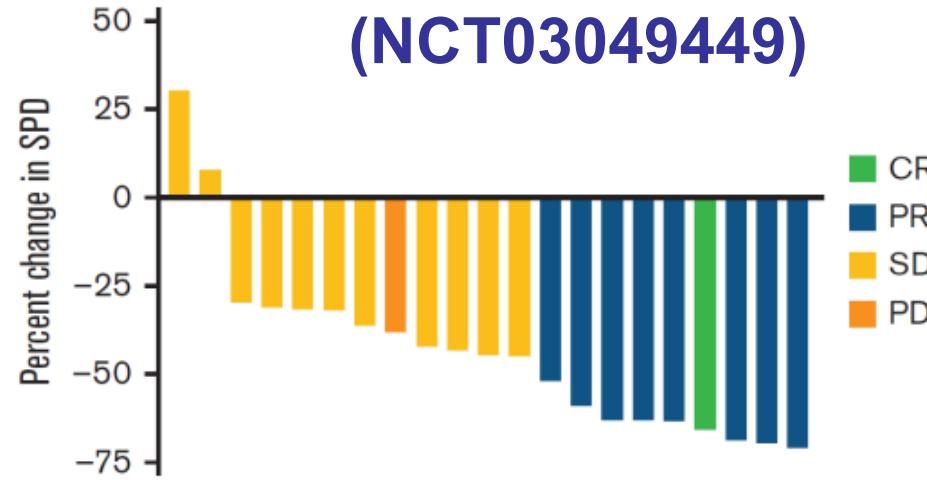
# Long term follow-up of CHARIOT trial

Response (data available for 12 pt)	N (%)
Objective response rate (ORR)	9 (75%)
Complete remission (CR)	6 (50%)
Partial response (PR)	3 (25%)
Median duration of response (range)	8.8 months (2.7–45.3 months)

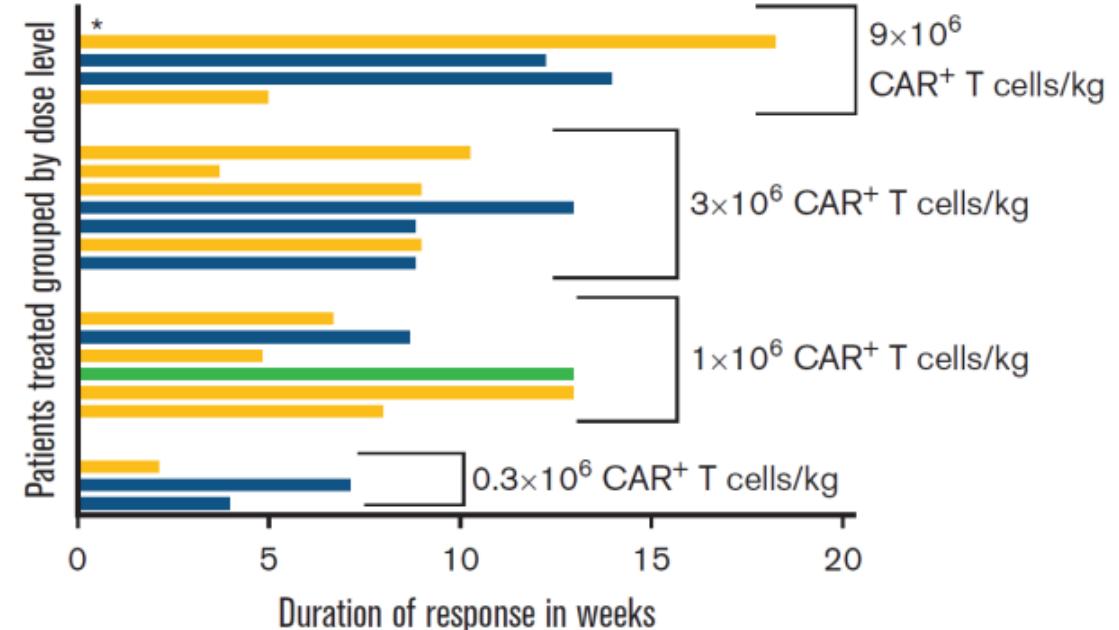
(Ahmed *et al.*, ASH 2025)



# Experience at NIH, Bethesda



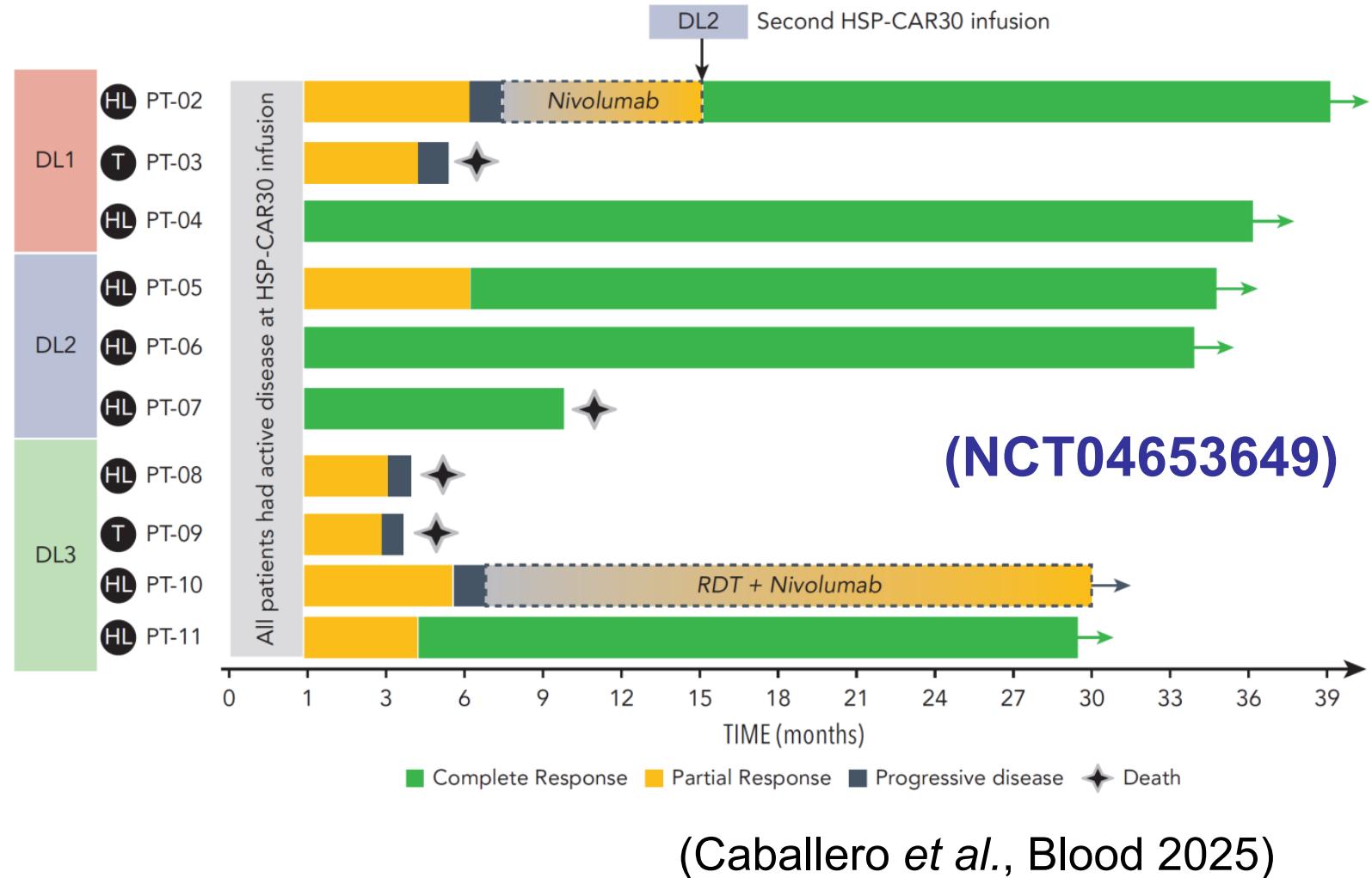
- 20 HL patients treated in phase 1 trial
- More toxicities, 2 dose-limiting:
  - 9 patients had rash, but 2 required prolonged steroid course
  - 5 had grade 3-4 cytopenias, with 2 complicated by life-threatening sepsis
- ORR 43%, CRR 5%, median DOR ~9 wks
- Further development was discontinued



(Brudno *et al.*, Blood Adv 2024)

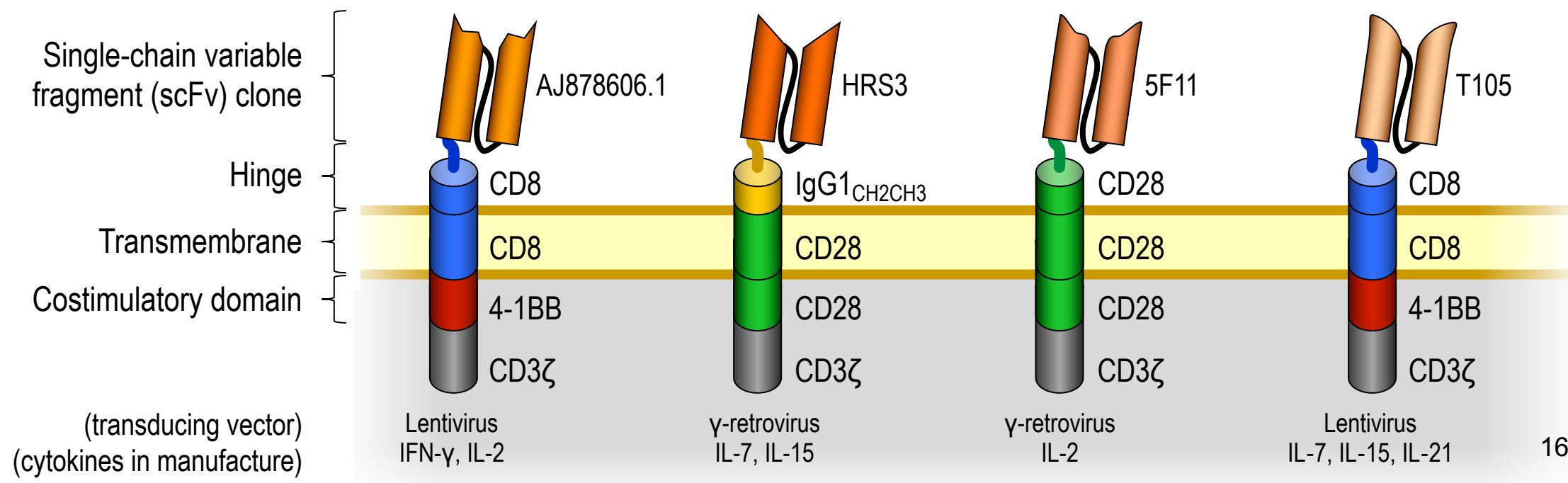
# Experience at H. Sant Pau, Barcelona

- 8 HL patients
  - Fresh product
  - Less “differentiated”
  - CARTs (IL-21 in culture)
  - More bendamustine
- Limited toxicity
- ORR 100%,  
CRR 63%
- All complete  
responses ongoing  
after a mean follow-  
up of 34 months



# Autologous CD30.CART studies in cHL

Study (year)	Wang et al. (2017)	Ramos et al. (2021)	Brudno et al. (2024)	Caballero et al. (2025)
Number of HL patients	17	42	20	8
Median age (range)	31 (13-55)	35 (17-69)	33 (18-64)	46 (21-63)
ORR   CR (%)	35   0	72   59	43   5	100   63
G3+ CRS   ICANS (%)	0   0	0   0	1   0	0   0
Other toxicities	Transient cytopenias	Transient rash Cytopenias	Rash requiring therapy Longer cytopenias	Transient rash Cytopenias, Infections



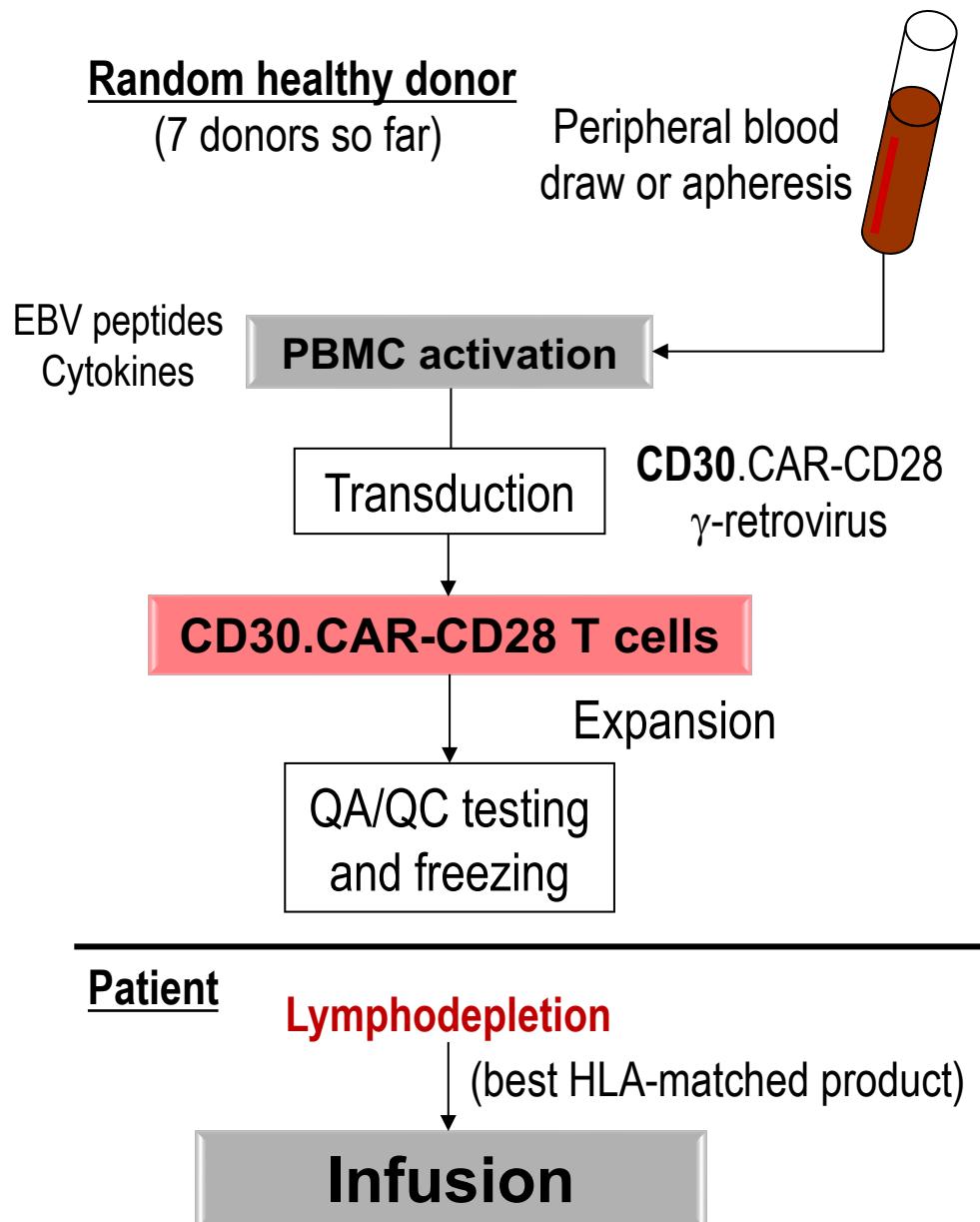
# Limitations of autologous CAR-T Cells

- Manufacture of individual patient-derived CAR T-cells
  - too time consuming to benefit acutely ill patients
  - prior chemotherapy exposure may result in suboptimal product
  - difficult to scale for large numbers of patients, expensive
- “Off-the-shelf” immune effector products that are banked from healthy donors would improve accessibility, allow rapid treatment, and reduce costs
  - need to avoid consequences of alloreactivity
    - Graft-versus-host disease (GVHD) and CAR-T cell rejection

# Why allogeneic CD30.CAR-EBVSTs?

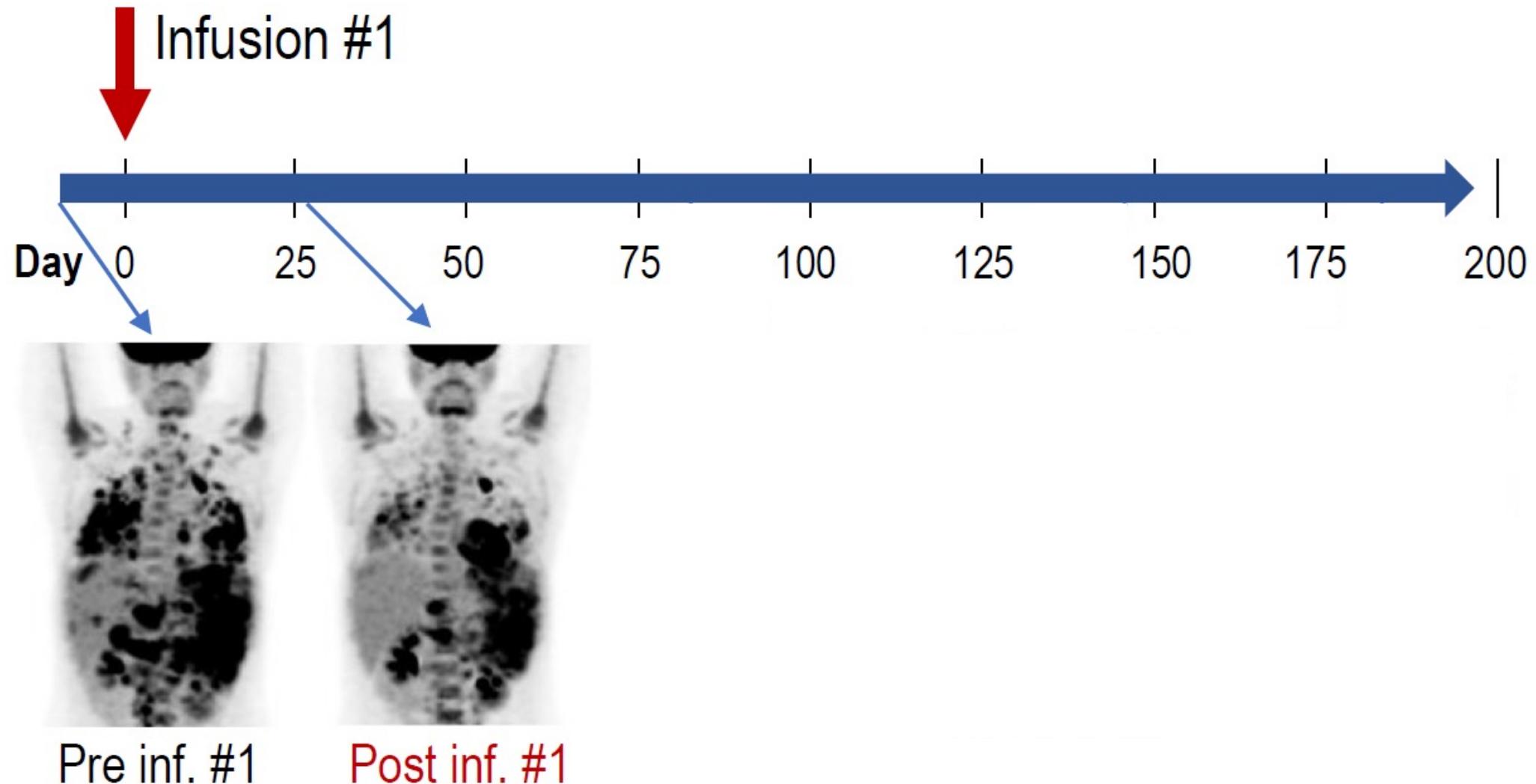
- Allogeneic EBV-specific T cells (EBVSTs) are safe in SCT and non-SCT recipients (Heslop, Sharma, Rooney, JCO 2021)
  - Manufactured from healthy individuals
  - Many patients treated in several trials without GVHD
  - Can localize to lymphoid tissues and sites of inflammation, proliferate *in vivo* and have potential to persist
- Activated T cells express CD30
  - Recipient T cells reacting against donor CAR-T cells may be killed by CD30.CAR-T cells
- May avoid GVHD and be protected from rejection

# BESTA clinical trial (NCT04288726)



- 26 enrollments (23 patients)
- Gender
  - 10 F, 13 M
- Diagnoses
  - Hodgkin lymphoma (21)
    - Nodular sclerosis (19)
    - Mixed cellularity (2)
  - Composite/gray zone lymphoma (2)
- Median age 35 yrs (range 22-62)
- Median 5 prior treatments (range 3-8)
  - PD-1 inhibitor (21), brentuximab vedotin (23), HDT/ASCT (12), allo-SCT (2), CD30.CAR-EBVST (3)

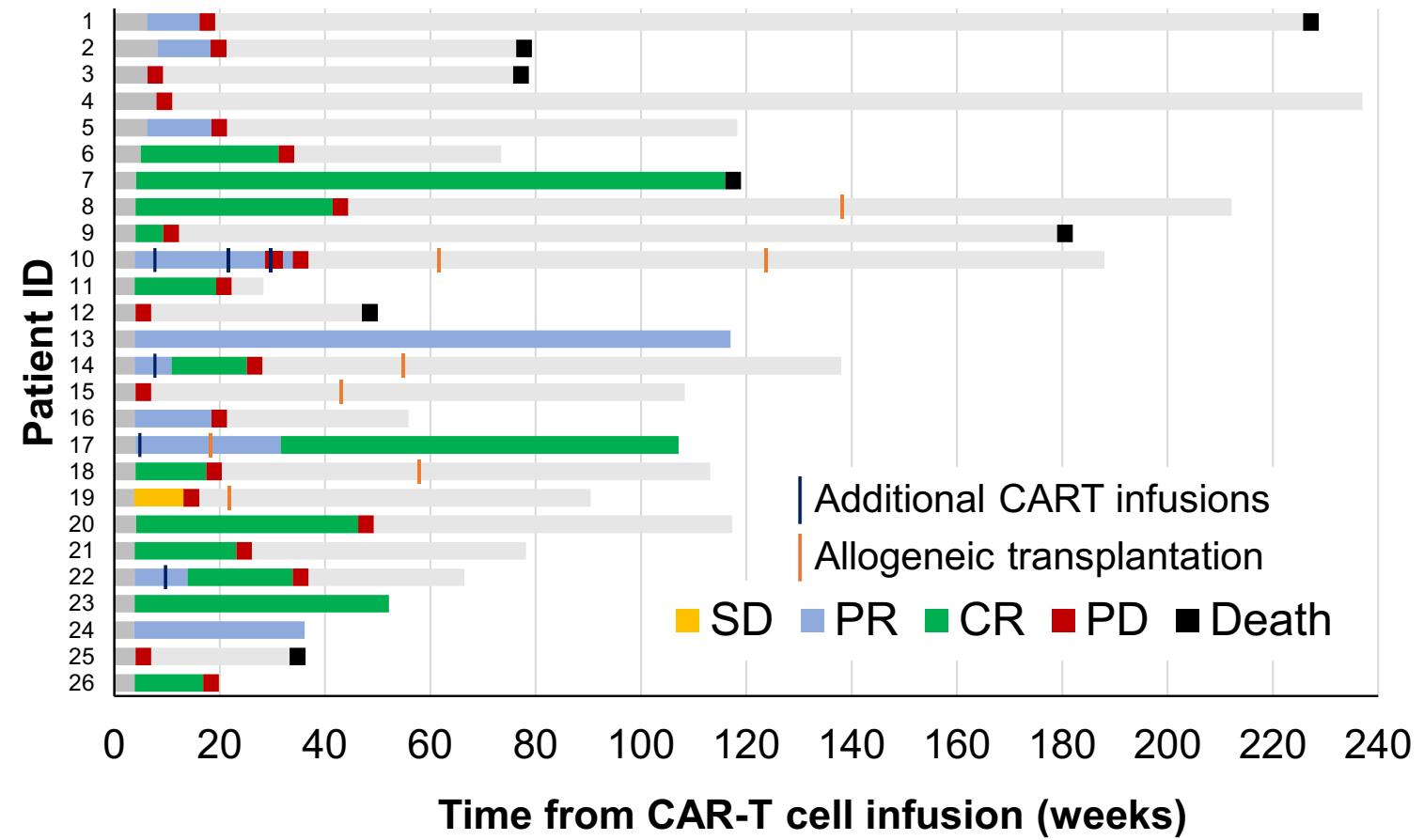
# Response to allogeneic CD30.CARTs



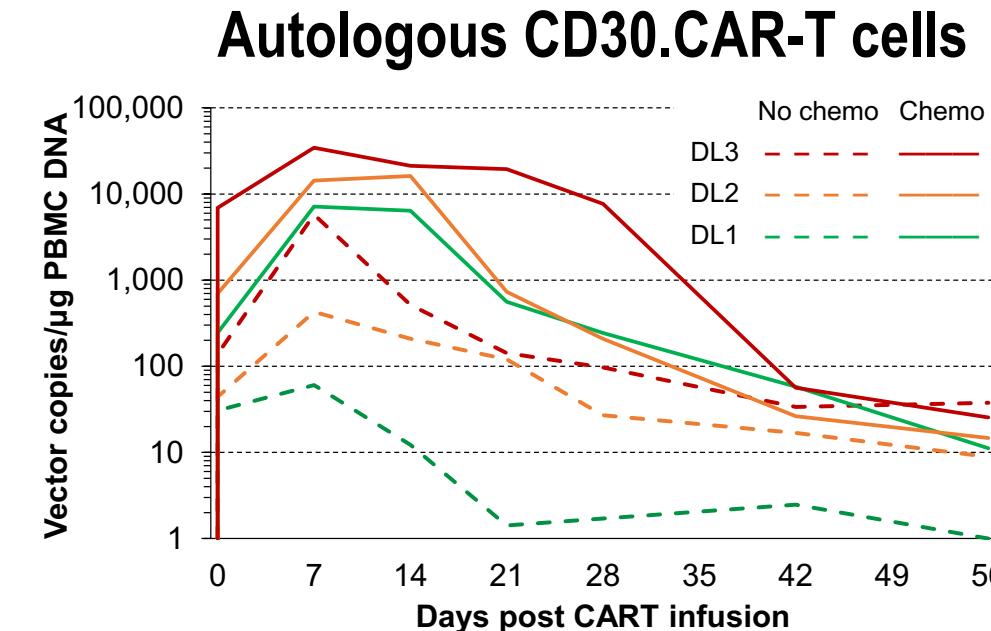
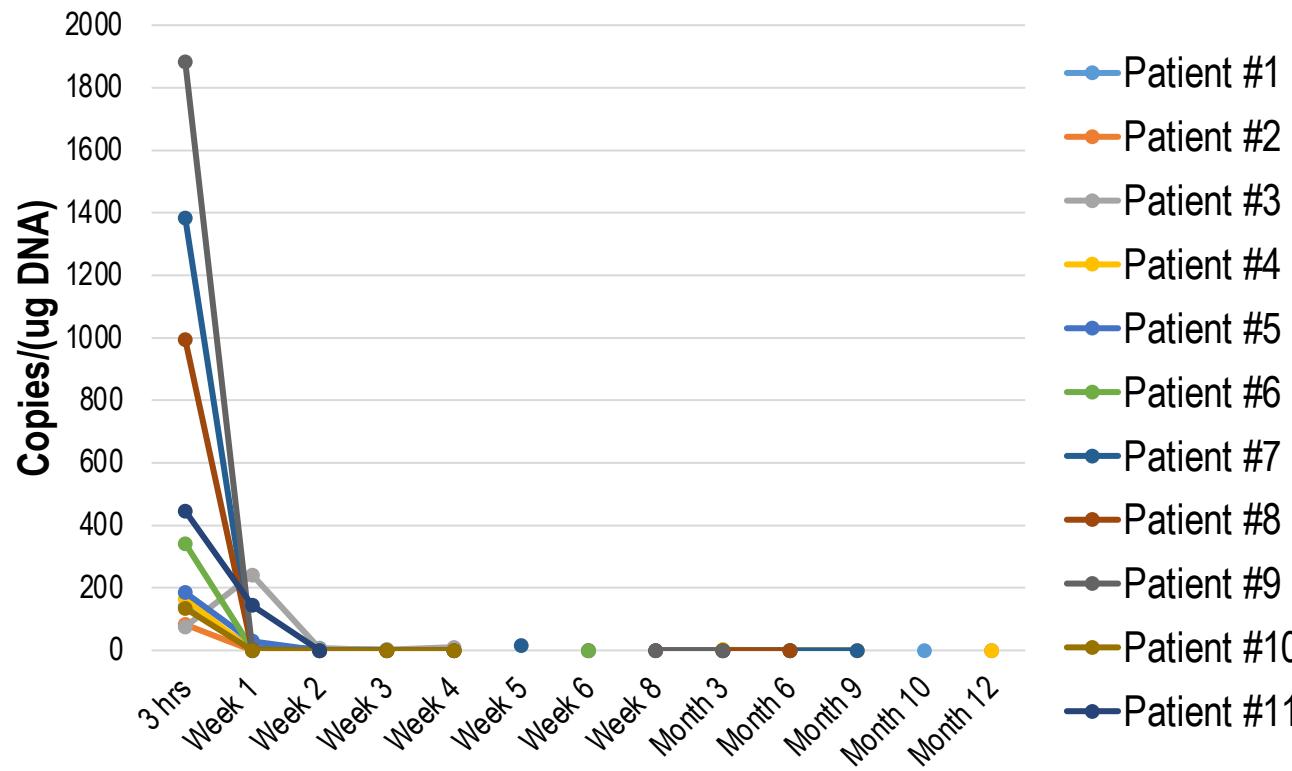
# Allo CD30.CAR-EBVST safety & response data

- No GVHD
  - Median 2 HLA matches
  - Range 1-7
- 9 episodes of CRS
  - All grade 1
- No ICANS
- Other AEs:
  - Mostly cytopenia due to chemo
    - 2 prolonged thrombocytopenias

77% ORR (20/26), 46% CR (12/26)



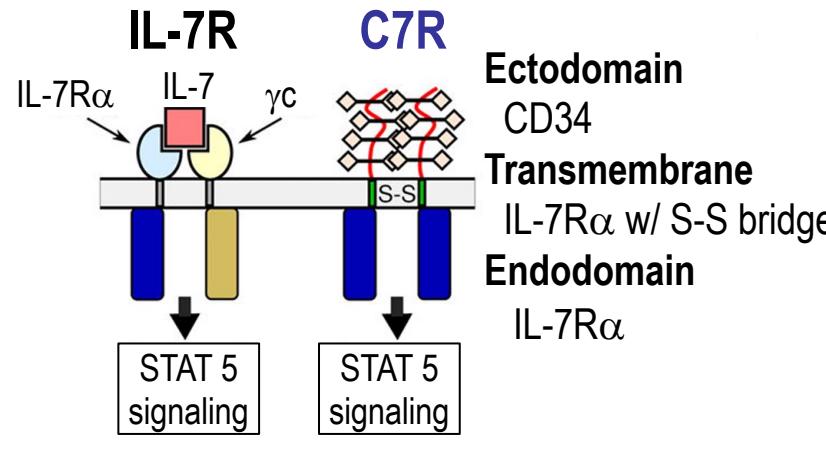
# Allogeneic CD30.CAR-EBVSTs have limited persistence in peripheral blood



Ramos CA, et al. JCI (2017) & JCO (2020)

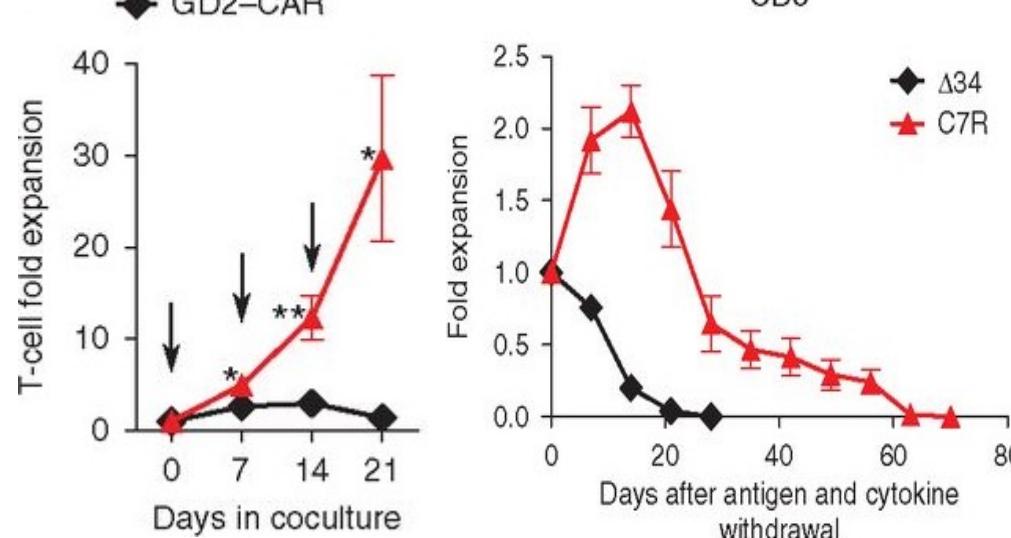
- Most patients show rapid loss of CD30.CAR EBVSTs in blood compared to autologous CD30.CAR-T cells with median DOR of ~24 vs 44 weeks
- Strategies to improve persistence are being developed:
  - E.g., constitutively active IL7 receptor expression in EBVSTs

# Constitutive IL-7R in CD30.CAR-EBVSTs



(wild type)

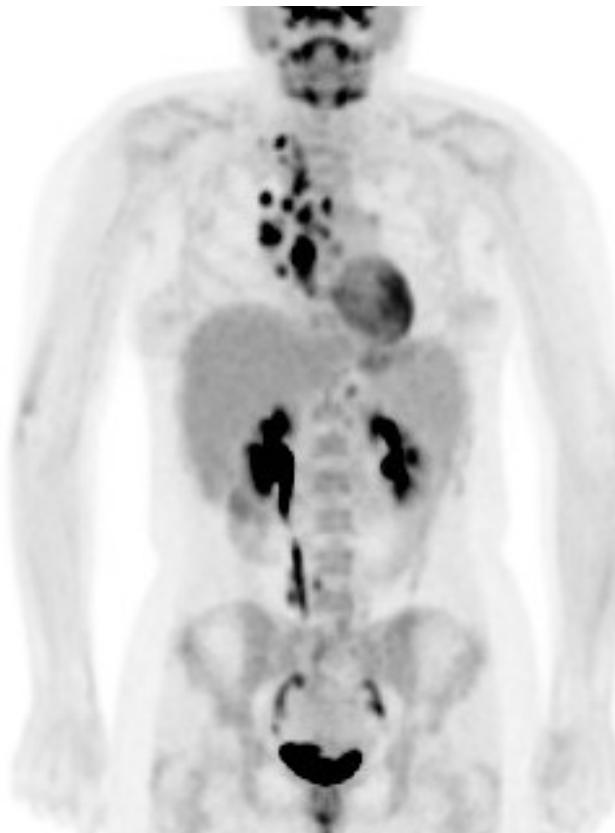
★ GD2-CAR.C7R  
◆ GD2-CAR



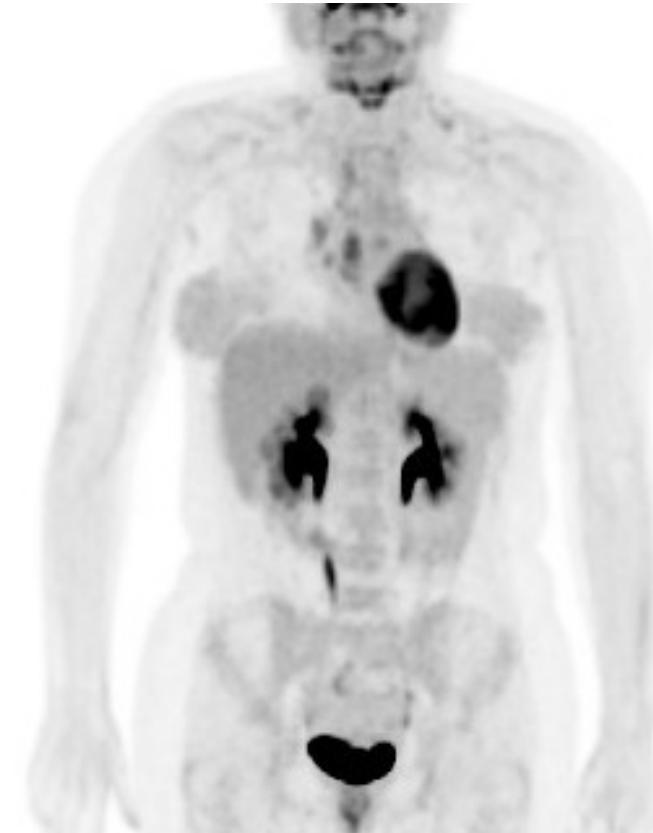
(Shum *et al.*, Cancer Discov 2017)

## CABAL2 trial (NCT06176690)

Pre-infusion



6 wks post-infusion



# Conclusions

- Adoptive transfer of autologous and allogeneic CD30.CARTs is feasible, safe, and potentially clinically effective
  - CRS and ICANS limited; maculopapular rash is seen often
- However, overall results are worse than those seen with autologous CD19.CART in NHL
  - But some products may be associated with better activity
- Allogeneic CD30.CAR EBVSTs lack persistence in patient blood
  - But immediate rejection does not seem be a major limitation
- Additional strategies are being explored to improve these results
  - But more patients and longer follow-up will be needed for validation
- But are we catching up? Unclear...
  - Cellular immune therapy seems to work for HL but...
  - Industry? Academia? Hybrid model?

# Grazie!

Cliona Rooney  
 Malcolm Brenner  
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David Quach  
 Haran Ganesh  
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 Sandhya Sharma  
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 Yezan Hadidi  
 Emily Hsieh

Sairah Ahmed  
 Jinwen Cao  
 Matthew Mei  
 Amer Beitinjaneh  
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 Rammurti Kamble  
 Vicky Torrano  
 Anaid Reyes  
 Bambi Grilley

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 Jessie Wu

**GMP Laboratories**  
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 Ka Liu

**GLP Laboratories**  
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**QA**  
 Natasha  
 Lapteva  
 Sara Richman

**QC**  
 Debbie Lyon

**All patients  
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